

Winter 12-5-2004

Chloride equilibrium potential in salamander cones.

Wallace B. Thoreson

University of Nebraska Medical Center, wbthores@unmc.edu

Eric J. Bryson

University of Nebraska Medical Center

Tell us how you used this information in this [short survey](#).

Follow this and additional works at: https://digitalcommons.unmc.edu/com_eye_articles

 Part of the [Ophthalmology Commons](#)

Recommended Citation

Thoreson, Wallace B. and Bryson, Eric J., "Chloride equilibrium potential in salamander cones." (2004).
Journal Articles: Ophthalmology. 54.
https://digitalcommons.unmc.edu/com_eye_articles/54

This Article is brought to you for free and open access by the Ophthalmology at DigitalCommons@UNMC. It has been accepted for inclusion in Journal Articles: Ophthalmology by an authorized administrator of DigitalCommons@UNMC. For more information, please contact digitalcommons@unmc.edu.

Research article

Open Access

Chloride equilibrium potential in salamander cones

Wallace B Thoreson* and Eric J Bryson

Address: Departments of Ophthalmology and Pharmacology, University of Nebraska Medical Center, Omaha, NE, USA

Email: Wallace B Thoreson* - walthores@unmc.edu; Eric J Bryson - ebryson@unmc.edu

* Corresponding author

Published: 05 December 2004

Received: 16 September 2004

BMC Neuroscience 2004, 5:53 doi:10.1186/1471-2202-5-53

Accepted: 05 December 2004

This article is available from: <http://www.biomedcentral.com/1471-2202/5/53>

© 2004 Thoreson and Bryson; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: GABAergic inhibition and effects of intracellular chloride ions on calcium channel activity have been proposed to regulate neurotransmission from photoreceptors. To assess the impact of these and other chloride-dependent mechanisms on release from cones, the chloride equilibrium potential (E_{Cl}) was determined in red-sensitive, large single cones from the tiger salamander retinal slice.

Results: Whole cell recordings were done using gramicidin perforated patch techniques to maintain endogenous Cl^- levels. Membrane potentials were corrected for liquid junction potentials. Cone resting potentials were found to average -46 mV. To measure E_{Cl} , we applied long depolarizing steps to activate the calcium-activated chloride current ($I_{Cl(Ca)}$) and then determined the reversal potential for the current component that was inhibited by the Cl^- channel blocker, niflumic acid. With this method, E_{Cl} was found to average -46 mV. In a complementary approach, we used a Cl^- -sensitive dye, MEQ, to measure the Cl^- flux produced by depolarization with elevated concentrations of K^+ . The membrane potentials produced by the various high K^+ solutions were measured in separate current clamp experiments. Consistent with electrophysiological experiments, MEQ fluorescence measurements indicated that E_{Cl} was below -36 mV.

Conclusions: The results of this study indicate that E_{Cl} is close to the dark resting potential. This will minimize the impact of chloride-dependent presynaptic mechanisms in cone terminals involving GABA_A receptors, glutamate transporters and $I_{Cl(Ca)}$.

Background

Regulation of intracellular chloride levels results in a chloride equilibrium potential (E_{Cl}) that is hyperpolarized with respect to the resting potential in many nerve cells, but depolarized in others [1-5]. For example, E_{Cl} in salamander rod photoreceptors is 25 mV more positive than the dark resting potential [6]. The resting potential of cone photoreceptors in darkness is around -42 to -47 mV and estimates of E_{Cl} in cones have ranged from -65 mV to -36 mV [7-11]. Cone photoreceptors possess a number of Cl^- conductances that help to shape their responses and syn-

aptic output. As discussed below, the value of E_{Cl} in cones is an important parameter for determining the strength and polarity of these effects.

It has been suggested GABA_A receptors in the terminals of cones may mediate inhibitory synaptic feedback from horizontal cells to cones [8]. Under this hypothesis, the light-evoked hyperpolarization of horizontal cells causes a cessation of GABA release and this disinhibition leads to a "feedback depolarization" in cones. There is evidence both for [e.g., [8]] and against [e.g., [12,13]; see review in

ref. [14])) this hypothesis. However, one prediction of the hypothesis is that the Cl^- equilibrium potential (E_{Cl}) must be negative to the resting potential in order for GABA disinhibition to depolarize a cone.

Cones possess prominent Ca^{2+} -activated Cl^- currents ($I_{\text{Cl}(\text{Ca})}$) [15-17] activated by the influx of Ca^{2+} through voltage-gated Ca^{2+} channels as well as by release of Ca^{2+} from intracellular stores [16]. Cl^- flux through $I_{\text{Cl}(\text{Ca})}$ can be substantial: during a 1.4 sec depolarizing step, the charge movement accompanying activation of $I_{\text{Cl}(\text{Ca})}$ is estimated to be 8.5 times that produced by activation of I_{Ca} alone [16]. These large membrane currents can strongly influence photoreceptor responses, but the nature of these effects depends on the value of E_{Cl} . If E_{Cl} is positive to the resting potential, activation of $I_{\text{Cl}(\text{Ca})}$ can boost depolarizing feedback responses from horizontal cells onto cones and produce prolonged, regenerative depolarizing responses lasting many seconds [9,18,19]. On the other hand, if E_{Cl} is negative to the resting potential, activation of $I_{\text{Cl}(\text{Ca})}$ can operate as a negative feedback mechanism to limit regenerative activation of Ca^{2+} channels [15,17]. In addition to altering membrane potential, depletion of intracellular Cl^- can directly inhibit the open channel probability of single Ca^{2+} channels, presumably by modifying an anion binding site on the intracellular surface of the channel [11]. In rods, where E_{Cl} is positive to the resting potential, there is evidence for a negative feedback pathway between I_{Ca} and $I_{\text{Cl}(\text{Ca})}$ in which activation of I_{Ca} stimulates $I_{\text{Cl}(\text{Ca})}$ leading to a Cl^- efflux that in turn inhibits Ca^{2+} channel activation [6,20]. If, however, E_{Cl} in cones is negative to the membrane potential, then activation of $I_{\text{Cl}(\text{Ca})}$ would stimulate an influx of Cl^- that would be expected to enhance Ca^{2+} channel open probability [11].

Cone photoreceptors have presynaptic glutamate transporters that are coupled to Cl^- channels [21-23]. The transporters in cones have been shown to respond to glutamate released from their own terminals [24]. Whether synaptically released glutamate causes cones to hyperpolarize or depolarize depends on E_{Cl} . Furthermore, analogous to the negative feedback from $I_{\text{Cl}(\text{Ca})}$ onto I_{Ca} described above, the chloride current produced by activation of glutamate transporters in rods can cause a Cl^- efflux that inhibits I_{Ca} [25]. As with the feedback between $I_{\text{Cl}(\text{Ca})}$ and I_{Ca} , the strength and polarity of this potential interaction in cones depends on E_{Cl} .

Given the importance of E_{Cl} in determining the impact of various feedback mechanisms in the photoreceptor terminal, we determined E_{Cl} in cone photoreceptors of the salamander retina using a combination of imaging with a chloride-sensitive dye and electrophysiological approaches.

Results

In control superfusate, dark resting potentials of cones from slices prepared under visible light averaged -46.0 ± 2.00 mV ($n = 9$) after correcting for the liquid junction potential. This is nearly identical to the dark resting potentials of salamander cones prepared under infrared illumination (-46.8 ± 2.03 mV, $n = 18$).

To measure E_{Cl} , $I_{\text{Cl}(\text{Ca})}$ was recorded using gramicidin perforated patch whole cell recordings and activated by applying a 500 ms step from -78 to -8 mV. This depolarizing step typically evoked a sustained inward tail current arising largely from activation of $I_{\text{Cl}(\text{Ca})}$ [19]. Only cells that exhibited an inward tail current were used for analysis. As shown in the example of Fig. 1A, the current/voltage relationship of a cone cell was assessed during the tail current by using a ramp voltage protocol (1 mV/ms from -98 to $+52$ mV) begun 25 ms after the end of the depolarizing step. The same protocol was then repeated after applying niflumic acid (0.1 mM; Fig. 1B). At this concentration, niflumic acid is a selective inhibitor of $I_{\text{Cl}(\text{Ca})}$ in cones [[19]; niflumic acid may not be as selective in rods: [20,26]]. Subtracting the control ramp-evoked current from that obtained in the presence of niflumic acid yields the current/voltage profile for $I_{\text{Cl}(\text{Ca})}$ (Fig. 1C). In the example shown in Fig. 1C, the difference current reversed around -46 mV. The reversal potential of the niflumic acid-sensitive difference current determined from 8 cones averaged -45.5 ± 2.5 mV. As a control for the possible perturbation of intracellular Cl^- by possible patch rupture, we repeated the same experiment using a pipette solution with only 3.5 mM Cl^- . E_{Cl} was not significantly different when measured using the low Cl^- pipette solution (-50.4 mV ± 3.4 mV; $n = 7$; $p = 0.49$, unpaired t-test). If patch rupture had occurred, E_{Cl} would be expected to attain -89 mV with the low Cl^- pipette solution and -20 mV with the original pipette solution.

Bath application of GABA evoked small reversible inward currents that averaged -3.4 ± 0.3 pA at the holding potential of -78 mV (not shown). The small size of these currents may be due to receptor desensitization [27]. Consistent with results obtained from measurements of $I_{\text{Cl}(\text{Ca})}$, difference currents calculated from ramps applied before and during GABA application indicate that the GABA-evoked current reversed at -46.4 ± 2.7 mV ($n = 9$).

In a complementary approach for measuring E_{Cl} , we used a Cl^- -sensitive dye, MEQ, to examine the Cl^- flux that accompanied cone depolarization evoked by bath application of various high K^+ solutions (12, 22, 31, 41, 50 and 70 mM K^+). In a separate set of experiments, we used gramicidin-perforated patch recording methods to measure the membrane potentials produced in cones by application of the different high K^+ solutions. Slices used for

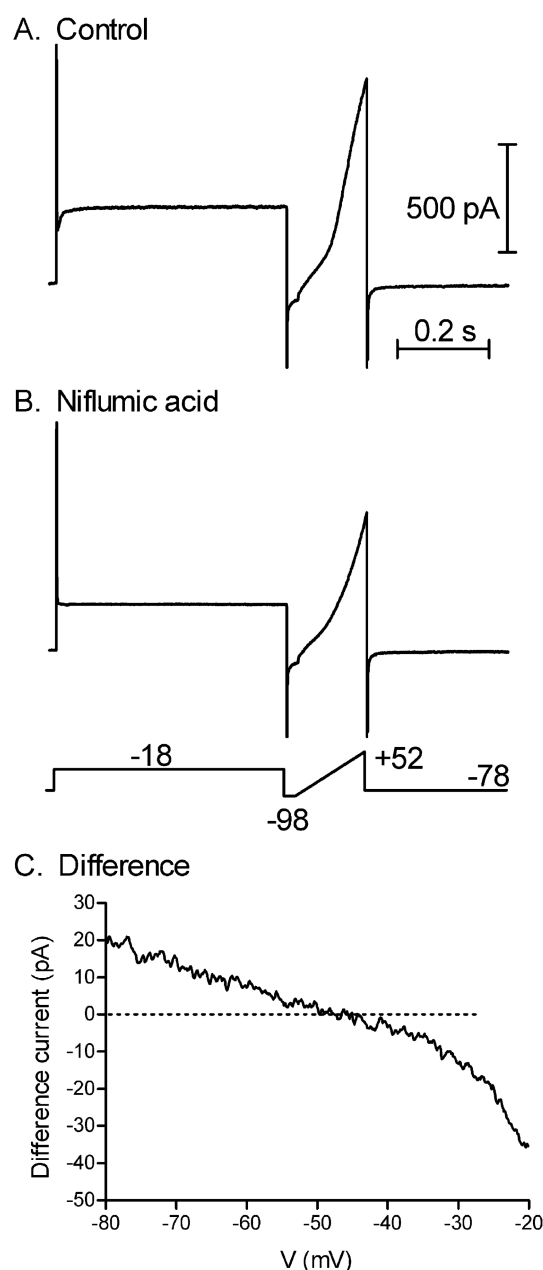


Figure 1

Cone E_{Cl} estimated from the reversal of $I_{Cl(Ca)}$. A) $I_{Cl(Ca)}$ tail current was activated by applying a 500 ms step from -78 to -18 mV during a gramicidin perforated patch whole cell recording from a rod. A ramp voltage protocol (-98 to +52 mV, 1 mV/ms) was applied during the tail current and begun 25 ms after termination of the step. B. The same protocol was then repeated in the presence of niflumic acid (0.1 mM) to inhibit $I_{Cl(Ca)}$. C. The ramp current/voltage relationship obtained in control medium (A) was subtracted from that obtained in the presence of niflumic acid (B) to yield a niflumic acid-sensitive difference current that reversed in this cell around -46 mV (after correcting for a liquid junction potential of -8 mV).

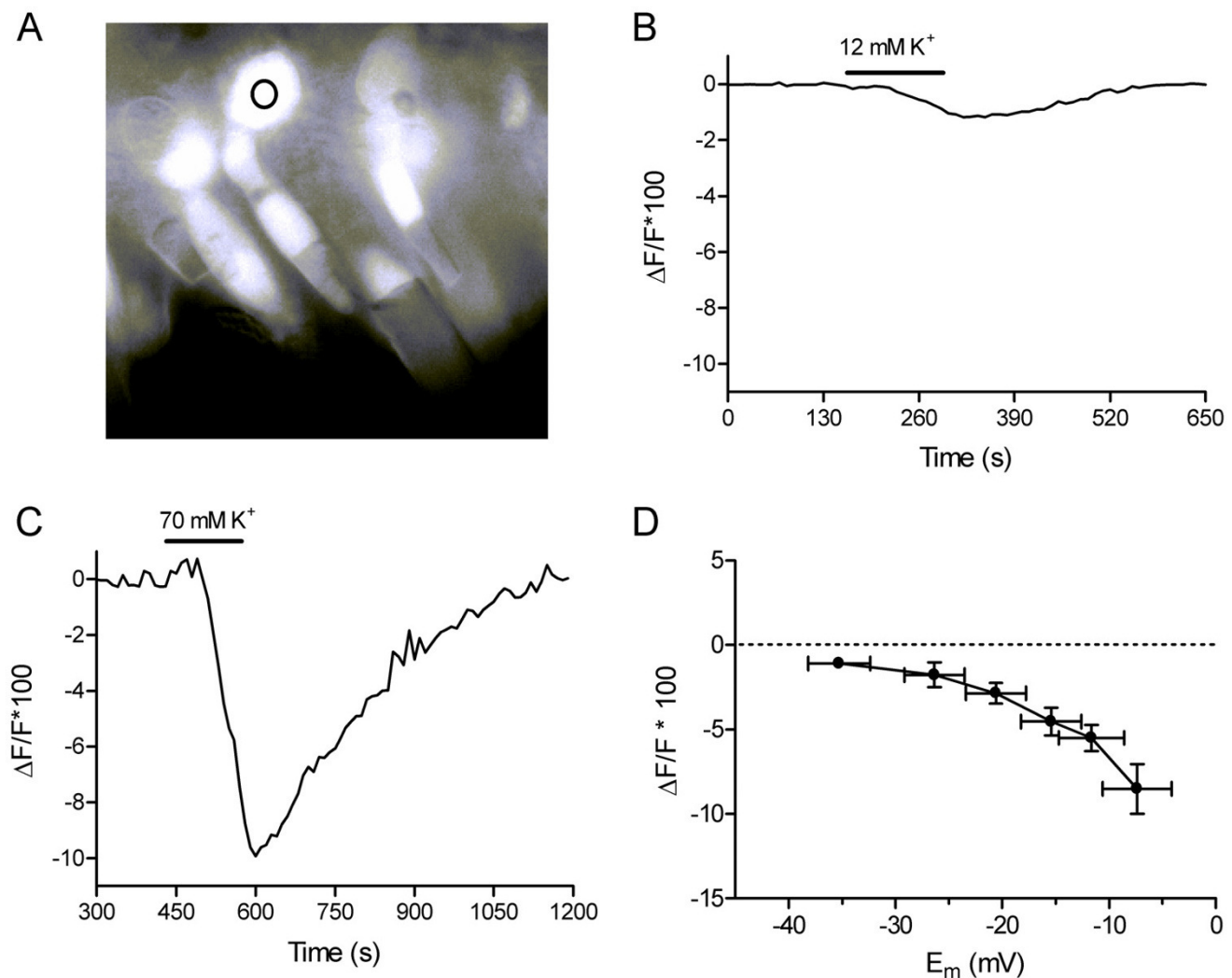
MEQ experiments and for measurement of membrane potentials in different solutions were prepared using similar techniques under visible illumination; control experiments showed that the fluorescent illumination used during MEQ experiments did not produce any further changes in the cone resting membrane potential ($n = 3$). An example of a retinal slice loaded with MEQ is shown in Fig. 2A. Measurements of MEQ fluorescence were made from the cone soma (circle, Fig. 2A). For a single wavelength dye such as MEQ, the change in fluorescence relative to basal fluorescence ($\Delta F/F$) can be used as a measure of the change in ion concentration [28]. In the cone in Fig. 2B, bath application of 12 mM K^+ , which depolarized cones to -36 mV, produced a 1.2% decrease in MEQ fluorescence. Since MEQ fluorescence is quenched by Cl^- ions this indicates that depolarization to -36 mV stimulated an influx of Cl^- ions. Application of a solution with 70 mM K^+ , which depolarizes cones to -7 mV, produced a greater influx of Cl^- as evidenced by the 10% decrease in MEQ fluorescence seen in a different cone (Fig. 2C). Fig. 2D shows the average change in $\Delta F/F$ ($\times 100$) plotted as a function of the membrane potential evoked by the different high K^+ solutions. The finding that 12 mM K^+ consistently stimulated an influx of Cl^- indicates that the reversal potential must be below -36 mV.

Discussion

The main finding of this study is that E_{Cl} in salamander cones is close to the dark resting potential (~ -46 mV). E_{Cl} was found to be -46 mV from block of $I_{Cl(Ca)}$ by niflumic acid; small GABA-evoked currents reversed around the same potential. MEQ fluorescence changes produced by depolarization support these electrophysiological measurements by indicating that E_{Cl} is below -36 mV.

There can be local variations of E_{Cl} within cells [4]. Large single cones in the salamander retina do not have a distinct axon and terminal; synaptic proteins are instead located at the base of the soma [29]. MEQ measurements were made in the cell soma from a region adjacent to the synaptic ending (see Fig. 2). $I_{Cl(Ca)}$ is localized to the terminal region in rods [30] and these channels are probably also localized to the terminals of cones. Thus, the measurements in the present study are likely to provide estimates of E_{Cl} in the synaptic terminal and adjacent regions of the cone cell. Measurements of intracellular Cl^- levels suggest that E_{Cl} in the inner segment is not significantly different from that measured in the soma [11].

The finding that the Cl^- equilibrium potential is close to the resting potential does not necessarily mean that Cl^- is passively distributed. Electrophysiological experiments required that cells be voltage clamped at -70 mV for many minutes. Nonetheless, the value of E_{Cl} determined from these electrophysiological experiments in which cells were

**Figure 2**

Intracellular chloride measurements. A. Example of a retinal slice fluorescently stained with the Cl^- -sensitive dye MEQ. The circle indicates the somatic region of a cone in which MEQ measurements were made. B. An example of the reduction in MEQ fluorescence, indicating an increase in intracellular $[Cl^-]$, produced by application of 12.1 mM K^+ to a cone. C. A greater decrease in MEQ fluorescence was produced by application of 69.9 mM K^+ . Different cone from panel B. D. MEQ fluorescence changes produced by bath application of superfusate containing 12.1, 21.6, 31.2, 40.7, 50.3, or 69.9 mM $[K^+]$. After correcting for liquid junction potentials, these high K^+ solutions depolarized cones to -35.3 ± 2.90 , -26.3 ± 2.82 , -20.6 ± 2.82 , -15.4 ± 2.84 , -11.6 ± 3.05 , and -7.4 ± 3.25 mV ($n = 9$), respectively, determined in current clamp recordings using gramicidin-perforated patch recording techniques. The change in MEQ fluorescence relative to basal fluorescence ($\Delta F/F \cdot 100$) is plotted against the cone membrane potential determined with each high K^+ solution.

voltage clamped at -70 mV was similar to the value estimated from MEQ studies in which cells were not voltage clamped and thus at their resting membrane potential. Results from experiments on the prolonged depolarization in cones also suggest that E_{Cl} can be maintained indefinitely at a value above the membrane potential. The plateau phase of the prolonged depolarization, which

largely reflects $I_{Cl(Ca)}$ activation [9,19], could remain above the membrane potential established by an adapting background for hours [9]. The ability of cones to maintain E_{Cl} above the membrane potential may arise from activity of the Na/KCl cotransporter as shown in rods (20) as well as from other mechanisms (e.g., CLC-2) [2,31].

Comparisons with other studies

E_{Cl} in cones has been estimated in a number of previous studies. The most positive value for E_{Cl} of -36 mV comes from calibration of MEQ fluorescence levels to determine the resting intracellular Cl^- concentrations in cones isolated from the salamander retina [11]. However, these measurements showed a large variability (range of S.E.M.: -26.5 to -46.6 mV). The most negative estimate of E_{Cl} comes from a study by Attwell et al [7] showing that the sign-reversing pathway from rods to cones reversed around -65 mV. Based on the presumption that this pathway involved disinhibition of GABAergic inputs into cones, this study has been interpreted as suggesting that E_{Cl} is around -65 mV. However, more recent evidence questions whether the horizontal cell to cone feedback pathway thought to underlie this sign-reversing pathway from rods to cones is truly GABAergic [9,13,14]. Other studies have arrived at values for E_{Cl} similar to those found in the present study. 1) By examining the polarity of GABA-evoked currents after patch rupture with either 12 or 24 mM Cl^- in the recording pipette, Kaneko and Tachibana [8] estimated E_{Cl} to be around -47 mV in isolated turtle cones. 2) Based on the membrane potential attained by the plateau phase of the prolonged depolarization in turtle cones from the eyecup slice preparation, E_{Cl} was estimated to be at or slightly above the dark resting potential of -42 mV [after correction for a liquid junction potential of -2 mV; ref. [9]]. 3) In a single recording from a salamander cone obtained with a Cl^- -sensitive electrode, Miller and Dacheux [32] found that E_{Cl} was 2 mV more positive than the dark resting potential. 4) A slightly more negative value for E_{Cl} was found in ruptured patch recordings from goldfish cones by examining the voltage dependence of the $I_{Cl(Ca)}$ tail current [10]. By extrapolating measurements back to the time of patch rupture, Kraaij et al [10] concluded that E_{Cl} was ~-55 mV.

Functional implications

I_{Ca} in cones, like that of rods, can be inhibited by lowering extracellular Cl^- [33]. The inhibition of I_{Ca} produced by lowering extracellular Cl^- appears to result from a reduction in intracellular Cl^- which in turn causes a reduction in the open probability of single Ca^{2+} channels [11]. In rods, where E_{Cl} is positive to the resting potential, activation of Cl^- channels leads to a Cl^- efflux thereby producing an inhibition of Ca^{2+} channels [6,11,20]. The present results indicate that activation of Cl^- channels when the cell is at its resting potential would produce minimal changes in intracellular Cl^- in cones. Therefore, the feedback between I_{Ca} and $I_{Cl(Ca)}$ postulated for rod photoreceptors [6,20] would be expected to be minimal in cones in darkness.

Another implication of the finding that E_{Cl} is close to the dark resting potential is that the stimulation of Cl^- channels associated with glutamate transporters by glutamate

released from cone terminals [24] would tend to stabilize the cell membrane potential near the dark potential. In rods, the Cl^- efflux accompanying activation of glutamate transporters appears to contribute to a glutamate-mediated inhibition of I_{Ca} [25]. As with the feedback between I_{Ca} and $I_{Cl(Ca)}$ considered in the previous paragraph, the finding that E_{Cl} is near the resting potential leads to the prediction that in darkness there would be no Cl^- efflux accompanying glutamate transporter activation and therefore glutamate would not be expected to inhibit I_{Ca} .

Cones hyperpolarize to light, although with prolonged illumination the membrane potential recovers to near the dark resting potential. The impact of chloride-dependent negative feedback between $I_{Cl(Ca)}$ and I_{Ca} or the glutamate transporter chloride current and I_{Ca} would be expected to increase as a cone hyperpolarizes in response to light. By reducing glutamate release, these chloride-dependent negative feedback mechanisms might thus contribute to making post-synaptic responses more transient.

The finding that E_{Cl} is near the resting potential of cones indicates that GABAergic disinhibition near the dark potential should produce little membrane potential change. This result is inconsistent with the postulated role for GABA in generating the feedback depolarization [8] and supports other studies suggesting that GABA is not directly responsible for horizontal to cone feedback [9,13,14].

Conclusions

Electrophysiological measurements, supported by experiments using chloride-sensitive dyes, indicate that E_{Cl} in salamander cones is close to the dark resting membrane potential. By minimizing the trans-membrane flux of chloride, this will minimize the presynaptic impact of GABA_A receptors, $I_{Cl(Ca)}$, and glutamate transporter chloride channels.

Methods

Tissue preparation

E_{Cl} is positive to the resting potential of many neurons in the immature brain [5]. Based on their size, the neotenus tiger salamanders (*Ambystoma tigrinum*, 15–25 cm) used in these experiments are thought to be 2–7 years old out of a life span of ~12 years [34].

Salamanders were handled humanely in accordance with protocols approved by the Institutional Animal Care and Use Committee at the University of Nebraska Medical Center. Chilled salamanders were rapidly decapitated, an eye was enucleated, and the front of the eye was removed. The resulting eyecup was cut into three or four pieces and a single piece was placed vitreal surface down onto a piece of filter paper (2 × 5 mm, Millipore type AAWP, 0.8 μm

pores). After adhering to the filter paper, the retina was isolated under chilled amphibian superfusate and cut into 125 μm slices using a razor blade tissue chopper (Stoelting Co., Wood Dale, IL). The slices were rotated 90° to view the retinal layers when placed under a water immersion objective (60X, 1.0 NA) on an upright fixed stage microscope (EF 600, Nikon Inc., USA). Slices were prepared under visible light but recordings were performed in darkness. All experiments were done using red-sensitive large single cones selected by anatomical criteria [35].

Solutions and perfusion

Solutions were applied with a single-pass, gravity-feed perfusion system (1 ml/min). The normal amphibian superfusate contained (in mM): 111 NaCl, 2.5 KCl, 1.8 CaCl_2 , 0.5 MgCl_2 , 10 N-2-hydroxyethylpiperazine-N' 2-ethanesulfonic acid (HEPES), and 5 glucose (pH 7.8). The osmolarity was measured with a vapor pressure osmometer (Wescor, Logan, UT) and adjusted, if necessary, to 242 ± 5 mOsm. For high K^+ solutions, various quantities of NaCl were replaced with equimolar KCl. Niflumic acid was diluted (1:10,000) from DMSO stock solutions. Unless otherwise specified, chemicals were obtained from Sigma/Aldrich/RBI (St. Louis, MO).

Electrophysiology

Patch pipettes were pulled on a PP-830 vertical puller (Narishige USA, New York) from borosilicate glass pipettes (1.2 mm O.D., 0.95 mm I.D., with internal filament) and had tips of ~ 1 μm outer diameter with resistances of 10 to 15 M Ω . To maintain endogenous levels of intracellular Cl^- , we obtained perforated patch whole cell recordings using the cation channel, gramicidin [36]. Gramicidin was dissolved in ethanol (5 mg/ml) and then added to the pipette electrolyte solution to achieve a final concentration of 5 $\mu\text{g/ml}$. For current clamp measurements of membrane potentials, the pipette electrolyte solution contained (in mM): 54 KCl, 61.5 KCH_3SO_4 (Pfaltz and Bauer, Waterbury, CT), 3.5 NaCH_3SO_4 , 10 HEPES. The pH was adjusted to 7.2 with KOH. The liquid junction potential (LJP) of this solution was estimated to be -7 mV using the junction potential calculator of PClamp (Axon Instruments). Membrane potential values reported throughout this manuscript were corrected for the LJP. For experiments with niflumic acid or GABA, pipettes were typically filled with a solution containing (in mM): 54 CsCl, 61.5 CsCH_3SO_3 , 3.5 NaCH_3SO_4 , 10 HEPES (LJP = -8 mV). In some experiments, a low Cl^- pipette solution was used containing: 115.5 mM CsCH_3SO_3 , 3.5 NaCl, 10 HEPES (LJP = -10 mV). The pH of both solutions was adjusted to 7.2 with CsOH. The osmolarity of pipette solutions were also adjusted, if necessary, to 242 ± 5 mOsm. Recordings were made using an Axopatch 200B amplifier (Axon Instruments Inc., Union City, CA) and PClamp 8 software (Axon Instruments).

Cell input resistance calculated using a step from -70 to -90 mV averaged 695 ± 111 M Ω . Access resistance estimated from the peak of the capacitive transient averaged 30.7 ± 4.8 M Ω ($n = 24$).

Imaging experiments

Digital fluorescent images were obtained with a cooled CCD camera (SensiCam, Cooke Corp., Auburn Hills, MI). Axon Imaging Workbench (AIW 2.2, Axon Instruments Inc., Union City, CA) was used to control the camera, filter wheel, and image acquisition. Pixel binning (2×2) of the images was used to decrease acquisition time to ≤ 1 s. Images were acquired at 5 to 10 s intervals during experimental trials.

For measurements of $[\text{Cl}^-]_i$ we used the dye, 6-methoxy-N-ethylquinolinium iodide (MEQ, Molecular Probes, Eugene, OR) [37]. MEQ was loaded into cells after reducing it to DiH-MEQ by adding 30 μM sodium borohydride (100 μl) to MEQ (5 mg) under a continuous stream of nitrogen gas [38]. DiHMEQ enters cells during the incubation period (15 min) where it is oxidized and retained in the form of MEQ. Fluorescence emission decreases as Cl^- quenches MEQ. The slow exponential decay in MEQ fluorescence due to dye leakage and bleaching was determined from a 3 min. series of control measurements prior to drug application and subtracted before analysis [11,20].

Variance is reported as $\pm$ S.E.M.

Authors' contributions

WT conceived the study and drafted the manuscript. WT and EB participated in all aspects of the experiments but most recordings were performed by EB. Both authors have read and approved the manuscript.

Acknowledgments

Supported by the National Eye Institute (EY10542), Research to Prevent Blindness, Inc., the Gifford Foundation, and the Nebraska Lions Foundation. The authors thank Dr. Dwight Burkhardt for his helpful comments on the manuscript.

References

1. Alvarez-Leefmans FJ, Gamino SM, Giraldez F, Nogueron I: **Intracellular chloride regulation in amphibian dorsal root ganglion neurones studied with ion-selective microelectrodes.** *J Physiol* 1988, **406**:225-246.
2. Staley K, Smith R, Schaack J, Wilcox C, Jentsch TJ: **Alteration of GABA_A receptor function following gene transfer of the CLC-2 chloride channel.** *Neuron* 1996, **17**:543-551.
3. Reuter D, Zierold K, Schroder WH, Frings S: **A depolarizing chloride current contributes to chemoelectrical transduction in olfactory sensory neurons in situ.** *J Neurosci* 1998, **18**:6623-6630.
4. Vardi N, Zhang LL, Payne JA, Sterling P: **Evidence that different cation chloride cotransporters in retinal neurons allow opposite responses to GABA.** *J Neurosci* 2000, **20**:7657-7663.
5. Ben-Ari Y: **Excitatory actions of gaba during development: the nature of the nurture.** *Nature Rev Neurosci* 2002, **3**:728-739.

6. Thoreson WB, Stella SL Jr, Bryson EJ, Clements J, Witkovsky P: **D2-like dopamine receptors promote interactions between calcium and chloride channels that diminish rod synaptic transfer in the salamander retina.** *Vis Neurosci* 2002, **19**:235-247.
7. Attwell D, Werblin FS, Wilson M, Wu SM: **A sign-reversing pathway from rods to double and single cones in the retina of the tiger salamander.** *J Physiol* 1983, **336**:313-333.
8. Kaneko A, Tachibana M: **Effects of gamma-aminobutyric acid on isolated cone photoreceptors of the turtle retina.** *J Physiol* 1986, **373**:443-461.
9. Thoreson WB, Burkhardt DA: **Ionic influences on the prolonged depolarization of turtle cones in situ.** *J Neurophysiol* 1991, **65**:96-110.
10. Kraaij DA, Spekreijse H, Kamermans M: **The nature of surround-induced depolarizing responses in goldfish cones.** *J Gen Physiol* 2000, **115**:3-16.
11. Thoreson WB, Nitzan R, Miller RF: **Chloride efflux inhibits single calcium channel open probability in vertebrate photoreceptors: chloride imaging and cell-attached patch-clamp recordings.** *Vis Neurosci* 2000, **17**:197-206.
12. Thoreson WB, Burkhardt DA: **Effects of synaptic blocking agents on the depolarizing responses of turtle cones evoked by surround illumination.** *Vis Neurosci* 1990, **5**:571-583.
13. Verweij J, Kamermans M, Spekreijse H: **Horizontal cells feed back to cones by shifting the cone calcium-current activation range.** *Vision Res* 1996, **36**:3943-3993.
14. Kamermans M, Spekreijse H: **The feedback pathway from horizontal cells to cones. A mini review with a look ahead.** *Vision Res* 1999, **39**:2449-2468.
15. Maricq AV, Korenbrot JJ: **Calcium and calcium-dependent chloride currents generate action potentials in solitary cone photoreceptors.** *Neuron* 1988, **1**:503-515.
16. Barnes S, Hille B: **Ionic channels of the inner segment of tiger salamander cone photoreceptors.** *J Gen Physiol* 1989, **94**:719-743.
17. Yagi T, Macleish PR: **Ionic conductances of monkey solitary cone inner segments.** *J Neurophysiol* 1994, **71**:656-665.
18. Burkhardt DA, Gottesman J, Thoreson WB: **Prolonged depolarization in turtle cones evoked by current injection and stimulation of the receptive field surround.** *J Physiol* 1988, **407**:329-348.
19. Barnes S, Deschenes MC: **Contribution of Ca and Ca-activated Cl channels to regenerative depolarization and membrane bistability of cone photoreceptors.** *J Neurophysiol* 1992, **68**:745-755.
20. Thoreson WB, Bryson EJ, Rabl K: **Reciprocal interactions between calcium and chloride in rod photoreceptors.** *J Neurophysiol* 2003, **90**:1747-1753.
21. Eliasof S, Arriza JL, Leighton BH, Kavanaugh MP, Amara SG: **Excitatory amino acid transporters of the salamander retina: identification, localization, and function.** *J Neurosci* 1998, **18**:698-712.
22. Eliasof S, Werblin F: **Characterization of the glutamate transporter in retinal cones of the tiger salamander.** *J Neurosci* 1993, **13**:402-411.
23. Picaud SA, Larsson HP, Grant GB, Lecar H, Werblin F: **Glutamate-gated chloride channel with glutamate-transporter-like properties in cone photoreceptors of the tiger salamander.** *J Neurophysiol* 1995, **74**:1760-1771.
24. Picaud S, Larsson HP, Wellis DP, Lecar H, Werblin F: **Cone photoreceptors respond to their own glutamate release in the tiger salamander.** *Proc Natl Acad Sci USA* 1995, **92**:9417-9421.
25. Rabl K, Bryson EJ, Thoreson WB: **Activation of glutamate transporters in rods inhibits presynaptic calcium currents.** *Vis Neurosci* 2003, **20**:557-566.
26. Satoh TO, Yamada M: **Niflumic acid reduces the hyperpolarization-activated current (I(h)) in rod photoreceptor cells.** *Neurosci Res* 2001, **40**:375-381.
27. Epstein R, Grundfest H: **Desensitization of gamma aminobutyric acid (GABA) receptors in muscle fibers of the crab *Cancer borealis*.** *J Gen Physiol* 1970, **56**:33-45.
28. Helmchen F: **Calibration of fluorescent calcium indicators.** In: *Imaging Neurons: A Laboratory Manual* Edited by: Yuste R, Lanni F, Konnerth A. New York: Cold Spring Harbor Laboratory Press; 2000:32.1-32.9.
29. Sherry DM, Yang H, Standifer KM: **Vesicle-associated membrane protein isoforms in the tiger salamander retina.** *J Comp Neurol* 2001, **431**:424-436.
30. Macleish PR, Nurse CA: **Ion channel compartments in photoreceptors [abstract].** *Invest Ophthalmol Vis Sci* 2000, **41**:S494.
31. Bösl MR, Stein V, Hubner C, Zdebek AA, Jordt SE, Mukhopadhyay AK, Davidoff MS, Holstein AF, Jentsch TJ: **Male germ cells and photoreceptors, both dependent on close cell-cell interactions, degenerate upon CIC-2 Cl channel disruption.** *EMBO J* 2001, **20**:1289-1299.
32. Miller RF, Dacheux RF: **Intracellular chloride in retinal neurons: measurement and meaning.** *Vision Res* 1983, **23**:399-411.
33. Thoreson WB, Nitzan R, Miller RF: **Reducing extracellular Cl⁻ suppresses dihydropyridine-sensitive Ca²⁺ currents and synaptic transmission in amphibian photoreceptors.** *J Neurophysiol* 1997, **77**:2175-2190.
34. Townes-Anderson E, Colantonio A, St Jules RS: **Age-related changes in the tiger salamander retina.** *Exp Eye Res* 1998, **66**:653-667.
35. Stella SL Jr, Thoreson WB: **Differential modulation of rod and cone calcium currents in tiger salamander retina by D2 dopamine receptors and cAMP.** *Eur J Neurosci* 2000, **12**:3537-3548.
36. Kyzozis A, Reichling DB: **Perforated-patch recording with gramicidin avoids artifactual changes in intracellular chloride concentration.** *J Neurosci Methods* 1995, **57**:27-35.
37. Biwersi J, Verkman AS: **Cell-permeable fluorescent indicator for cytosolic chloride.** *Biochemistry* 1991, **30**:7879-7883.
38. Woll E, Gschwentner M, Furst J, Hofer S, Buemberger G, Jungwirth A, Frick J, Deetjen P, Paulmichl M: **Fluorescence-optical measurements of chloride movements in cells using the membrane-permeable dye diH-MEQ.** *Pflugers Arch* 1996, **432**:486-493.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

