

1-1-2011

## The incidence of type-1 diabetes in NOD mice is modulated by restricted flora not germ-free conditions.

Cecile King  
*Scripps Research Institute*

Nora Sarvetnick  
*University of Nebraska Medical Center, noras@unmc.edu*

Follow this and additional works at: [https://digitalcommons.unmc.edu/reg\\_articles](https://digitalcommons.unmc.edu/reg_articles)



Part of the [Molecular Biology Commons](#), and the [Molecular, Cellular, and Tissue Engineering Commons](#)

---

### Recommended Citation

King, Cecile and Sarvetnick, Nora, "The incidence of type-1 diabetes in NOD mice is modulated by restricted flora not germ-free conditions." (2011). *Journal Articles: Regenerative Medicine*. 19.  
[https://digitalcommons.unmc.edu/reg\\_articles/19](https://digitalcommons.unmc.edu/reg_articles/19)

This Article is brought to you for free and open access by the Regenerative Medicine at DigitalCommons@UNMC. It has been accepted for inclusion in Journal Articles: Regenerative Medicine by an authorized administrator of DigitalCommons@UNMC. For more information, please contact [digitalcommons@unmc.edu](https://digitalcommons.unmc.edu).

# The Incidence of Type-1 Diabetes in NOD Mice Is Modulated by Restricted Flora Not Germ-Free Conditions

Cecile King<sup>\*a</sup>, Nora Sarvetnick<sup>ab</sup>

Department of Immunology, The Scripps Research Institute, La Jolla, California, United States of America

## Abstract

In the NOD mouse, the incidence of type-1 diabetes is thought to be influenced by the degree of cleanliness of the mouse colony. Studies collectively demonstrate that exposure to bacterial antigen or infection in the neonatal period prevents diabetes [1,2,3,4,5,6,7,8,9,10], supporting the notion that immunostimulation can benefit the maturation of the postnatal immune system [11]. A widely accepted extrapolation from this data has been the notion that NOD mice maintained under germ-free conditions have an increased incidence of diabetes. However, evidence supporting this influential concept is surprisingly limited [12]. In this study, we demonstrate that the incidence of diabetes in female NOD mice remained unchanged under germ-free conditions. By contrast, a spontaneous monoculture with a gram-positive aerobic spore-forming rod delayed the onset and reduced the incidence of diabetes. These findings challenge the view that germ-free NOD mice have increased diabetes incidence and demonstrate that modulation of intestinal microbiota can prevent the development of type-1 diabetes.

**Citation:** King C, Sarvetnick N (2011) The Incidence of Type-1 Diabetes in NOD Mice Is Modulated by Restricted Flora Not Germ-Free Conditions. PLoS ONE 6(2): e17049. doi:10.1371/journal.pone.0017049

**Editor:** Matthias von Herrath, La Jolla Institute of Allergy and Immunology, United States of America

**Received:** October 18, 2010; **Accepted:** January 14, 2011; **Published:** February 25, 2011

**Copyright:** © 2011 King, Sarvetnick. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was supported by grants from the NIH/DK54063 and Juvenile Diabetes Research Foundation. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: c.king@garvan.org.au

<sup>a</sup> Current address: Department of Immunology, The Garvan Institute of Medical Research, Darlinghurst, New South Wales, Australia

<sup>b</sup> Current address: Lied Transplant Center, University of Nebraska Medical Center, Omaha, Nebraska, United States of America

## Introduction

The development of the postnatal immune system is guided by the interactions of lymphocytes with self-MHC/peptide ligands derived from our body's own tissues and those from the environment, such as the commensal microbial flora of the gastrointestinal tract and the diet. The important role of the gastrointestinal microbiota has been emphasized by the evidence of reduced intestinal lymphatic tissue and underdeveloped lymphoid organs in germ-free mice [13,14,15,16]. The interplay between the host immune system and commensal bacteria is dynamic and continuous since the size and cellularity of gastrointestinal lymphoid tissues recover following selective colonization of germ-free animals [13,14]. Studies on local immune function demonstrate that IgA secreting plasmablasts are reduced in germ free mice [17] and the induction of commensal specific IgA [18] has been shown to occur in response to current bacterial exposure [19]. The molecular basis for the influence of commensal bacteria on host immune function remains incompletely understood, but recent studies have revealed a critical role for the nucleotide-binding oligomerization domain containing 1 (NOD1) protein [20] and bacterial polysaccharide [21].

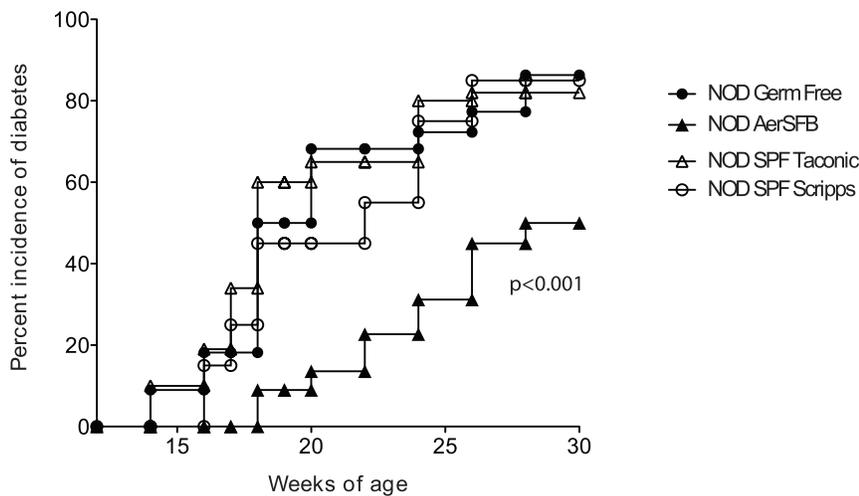
The way in which antigenic stimulation guides the development and maintenance of a healthy immune system is of fundamental importance to our understanding of immunological tolerance. In the non-obese diabetic (NOD) mouse strain, the target pancreatic insulin producing beta cells are attacked and destroyed by activated immune cells, leading to type-1 diabetes. Several infectious, and non-infectious agents are known to prevent type-1 diabetes in NOD mice. They include; persistent viral infection

(MHV [1], LCMV [2]), mycobacterial infection [3], bacterial antigens [4,5]; Hsp65 [6,7,8] and complete (heat-killed mycobacterium-containing) Freund's adjuvant (CFA) [9,10]. Stimulation with adjuvant containing bacterial extracts in the neonatal period is known to prevent diabetes and imparts qualitative and quantitative changes in the immune cell compartments that lasts throughout adulthood [9,22]. There has been a number of hypotheses presented that could account for the protective effect of immunostimulation such as a change in the cytokine milieu [23] and the increase in T cell numbers or populations of regulatory T cells [5,22,24,25,26].

Thus, the incidence of type-1 diabetes in NOD mice is thought to reflect the degree of cleanliness of the colony. A widely accepted extrapolation from these data has been that NOD mice maintained under germ-free conditions have an increased incidence of diabetes. However, there is little evidence to support this view [12].

## Materials and Methods

A germfree caesarean derivation was performed on NOD mice using 10 females and 5 males. Caesarean-derived mice were weaned onto sterile dams and housed in sterile isolators to maintain germfree status and fed an endotoxin free NIH31-M diet, designed to provide appropriate nutrition after being autoclaved <http://www.taconic.com/wmspage.cfm?parm1=292> until thirty weeks of age at Taconic Farms breeding facility, Germantown, NY, USA. All supplies were sterilized and entered into the isolator using strict aseptic techniques. Mice underwent monthly microbiological



**Figure 1. The incidence of type-1 diabetes in female NOD mice remains unchanged under germ-free conditions but is reduced by restricted flora.** Cumulative diabetes incidence of germ-free female NOD mice (NOD germ-free  $n = 22$ ), female NOD mice monocolonized with an aerobic spore-forming bacteria (*Bacillus cereus*) detected at week 16 (NOD AerSFB  $n = 22$ ) and female NOD mice housed under specific pathogen-free conditions (NOD SPF Scripps  $n = 22$ , NOD SPF Taconic  $n = 40$ ). Urine glucose was measured every one or two weeks as shown and mice that had reached 4+ (55 mmol/L) were considered diabetic. There was a significant inhibition ( $p < 0.001$ ) in the incidence of diabetes in monocolonized NOD mice compared with SPF NOD mice, but no significant difference between germ free and SPF NOD mice ( $p > 0.05$ , Anova). doi:10.1371/journal.pone.0017049.g001

testing to ensure that germfree rodents were free of aerobic and anaerobic organisms (for a complete list of agents see <http://www.taconic.com/wmspage.cfm?parm1=265>). Urine glucose was measured every one or two weeks and mice with frank glucosuria -two consecutive reading of 4+ (estimated at 55 mmol/L urine glucose) were considered diabetic. Control female NOD mice were housed under specific pathogen-free (SPF) conditions at Taconic and the Scripps rodent colony, which were handled in accordance with the TSRI Animal Care and Use Committee, which approved this study (A3194-01). Blood glucose values (BGV) were determined using Glucofilm blood glucose strips (Miles Diagnostic, Elkhart, IN). Mice were considered diabetic following two consecutive blood glucose readings above 18 mmol/L.

## Results and Discussion

We sought to determine the influence of intestinal microbiota on the incidence of type-1 diabetes in NOD mice. To our surprise, the incidence of type-1 diabetes in female germ free NOD mice ( $n = 22$ ) was indistinguishable from that of NOD mice housed under SPF conditions in our colony ( $n = 20$ ) (Figure 1,  $p > 0.6696$ ) or the 80% incidence of female NOD mice housed under SPF conditions at Taconic (Figure 1,  $p < 0.78$ ). However, our findings also indicated that intestinal microflora had the capacity to influence the development of type-1 diabetes. In one cohort of NOD mice ( $n = 22$ ) housed in a separate isolator, a spontaneous contamination with a gram-positive aerobic spore-forming rod (that was subsequently typed as *Bacillus cereus*) was detected at week 16. These mice exhibited a delayed onset, and reduced incidence of clinical disease ( $p < 0.001$ ) during the 30-week study period (Figure 1).

## References

- Wilberz S, Partke HJ, Dagnaes-Hansen F, Herberg L (1991) Persistent MHV (mouse hepatitis virus) infection reduces the incidence of diabetes mellitus in non-obese diabetic mice. *Diabetologia* 34: 2–5.
- Oldstone MB (1988) Prevention of type I diabetes in nonobese diabetic mice by virus infection. *Science* 239: 500–502.
- Martins TC, Aguas AP (1996) Changes in B and T lymphocytes associated with mycobacteria-induced protection of NOD mice from diabetes. *J Autoimmun* 9: 501–507.
- Satoh J, Shintani S, Oya K, Tanaka S, Nobunaga T, et al. (1988) Treatment with streptococcal preparation (OK-432) suppresses anti-islet autoimmunity and prevents diabetes in BB rats. *Diabetes* 37: 1188–1194.
- Qin HY, Singh B (1997) BCG vaccination prevents insulin-dependent diabetes mellitus (IDDM) in NOD mice after disease acceleration with cyclophosphamide. *J Autoimmun* 10: 271–278.
- Elias D, Markovits D, Reshef T, van der Zee R, Cohen IR (1990) Induction and therapy of autoimmune diabetes in the non-obese diabetic (NOD/Lt)

- mouse by a 65-kDa heat shock protein. *Proc Natl Acad Sci U S A* 87: 1576–1580.
7. Sai P, Rivereau AS (1996) Prevention of diabetes in the nonobese diabetic mouse by oral immunological treatments. Comparative efficiency of human insulin and two bacterial antigens, lipopolysaccharide from *Escherichia coli* and glycoprotein extract from *Klebsiella pneumoniae*. *Diabetes Metab* 22: 341–348.
  8. Ramiya VK, Shang XZ, Pharis PG, Wasserfall CH, Stabler TV, et al. (1996) Antigen based therapies to prevent diabetes in NOD mice. *J Autoimmun* 9: 349–356.
  9. Sadelain MW, Qin HY, Lauzon J, Singh B (1990) Prevention of type I diabetes in NOD mice by adjuvant immunotherapy. *Diabetes* 39: 583–589.
  10. McInerney MF, Pek SB, Thomas DW (1991) Prevention of insulinitis and diabetes onset by treatment with complete Freund's adjuvant in NOD mice. *Diabetes* 40: 715–725.
  11. Holt PG (1995) Postnatal maturation of immune competence during infancy and childhood. *Pediatr Allergy Immunol* 6: 59–70.
  12. Maclaren N, Schatz D, Drash A, Grave G (1989) Initial pathogenic events in IDDM. *Diabetes* 38: 534–538.
  13. Hudson JA, Luckey TD (1964) Bacteria Induced Morphologic Changes. *Proc Soc Exp Biol Med* 116: 628–631.
  14. Glaister JR (1973) Factors affecting the lymphoid cells in the small intestinal epithelium of the mouse. *Int Arch Allergy Appl Immunol* 45: 719–730.
  15. Hooijkaas H, Benner R, Pleasants JR, Wostmann BS (1984) Isotypes and specificities of immunoglobulins produced by germ-free mice fed chemically defined ultrafiltered "antigen-free" diet. *Eur J Immunol* 14: 1127–1130.
  16. Cebra JJ, Periwal SB, Lee G, Lee F, Shroff KE (1998) Development and maintenance of the gut-associated lymphoid tissue (GALT): the roles of enteric bacteria and viruses. *Dev Immunol* 6: 13–18.
  17. Crabbe PA, Nash DR, Bazin H, Eysen H, Heremans JF (1970) Immunohistochemical observations on lymphoid tissues from conventional and germ-free mice. *Lab Invest* 22: 448–457.
  18. Macpherson AJ, Gatto D, Sainsbury E, Harriman GR, Hengartner H, et al. (2000) A primitive T cell-independent mechanism of intestinal mucosal IgA responses to commensal bacteria. *Science* 288: 2222–2226.
  19. Hapfelmeier S, Lawson MA, Slack E, Kirundi JK, Stoel M, et al. (2010) Reversible microbial colonization of germ-free mice reveals the dynamics of IgA immune responses. *Science* 328: 1705–1709.
  20. Bouskra D, Brezillon C, Berard M, Werts C, Varona R, et al. (2008) Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. *Nature* 456: 507–510.
  21. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL (2005) An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 122: 107–118.
  22. King C, Ilic A, Koelsch K, Sarvetnick N (2004) Homeostatic Expansion of T cells during immune insufficiency Generates Autoimmunity. *Cell* 117: 265–277.
  23. Shehadeh NN, LaRosa F, Lafferty KJ (1993) Altered cytokine activity in adjuvant inhibition of autoimmune diabetes. *J Autoimmun* 6: 291–300.
  24. Gombert JM, Herbelin A, Tancrede-Bohin E, Dy M, Chatenoud L, et al. (1996) Early defect of immunoregulatory T cells in autoimmune diabetes. *C R Acad Sci III* 319: 125–129.
  25. Baxter AG, Kinder SJ, Hammond KJ, Scollay R, Godfrey DI (1997) Association between alpha $\beta$ TCR+CD4 $\supset$ CD8 $\supset$  T-cell deficiency and IDDM in NOD/Lt mice. *Diabetes* 46: 572–582.
  26. Martins TC, Aguas AP (1999) Mechanisms of *Mycobacterium avium*-induced resistance against insulin-dependent diabetes mellitus (IDDM) in non-obese diabetic (NOD) mice: role of Fas and Th1 cells. *Clin Exp Immunol* 115: 248–254.
  27. Elliott RB, Reddy SN, Bibby NJ, Kida K (1988) Dietary prevention of diabetes in the non-obese diabetic mouse. *Diabetologia* 31: 62–64.
  28. Mueller DB, Koczwara K, Mueller AS, Pallauf J, Ziegler AG, et al. (2009) Influence of early nutritional components on the development of murine autoimmune diabetes. *Ann Nutr Metab* 54: 208–217.
  29. Alam C, Valkonen S, Palagani V, Jalava J, Eerola E, et al. (2010) Inflammatory tendencies and overproduction of IL-17 in the colon of young NOD mice are counteracted with diet change. *Diabetes* 59: 2237–2246.
  30. Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, et al. (2008) Innate immunity and intestinal microbiota in the development of Type 1 diabetes. *Nature* 455: 1109–1113.